

Chemotherapy induced thrombocytopenia treated by four types of platelets concentrates

Ljubinka I. Nikolić¹, Ninoslav D. Nedeljković², Svetislav B. Jelić²,
Nada D. Suvajdžić Vuković³, Ivana M. Filipović-Lješković²,
Srdjan Z. Marković⁴, Drina Lj. Janković⁵, Dragana A. Kastratović⁴

¹ Clinic for Gynecology and Obstetrics, Clinical Centre of Serbia, Belgrade, Serbia

² Institute for Oncology and Radiology of Serbia, Belgrade, Serbia

³ Clinic for Hematology, Clinical Centre of Serbia, Belgrade, Serbia

⁴ Clinical Centre of Serbia, Belgrade, Serbia

⁵ Vinča Institute of Nuclear Sciences, Belgrade, Serbia

SUMMARY

Introduction: Serious adverse event of anticancer chemotherapy is granulocytopenia and thrombocytopenia which can decrease efficiency of final therapy results. After many years, platelet concentrates transfusion (PCT) is still researching problem without sure standpoint.

The aim: To determine whether there is a difference in the clinical efficiency in the use of 4 types of platelet applied for transfusion; - to ascertain whether platelet count increase expressed as corrected count increment (CCI), is a better parameter for the evaluation of platelet transfusion efficiency than the bleeding time (Bt), as the only readily assessable *in vivo* platelet function related parameter.

Subjects and methods: This paper is a part of academic (noncommercial) IV phase observational nonintervention study. Investigation included 78 patients diagnosed with malignant lymphoma and metastatic solid tumors, transfused by platelet concentrates. Patients were divided into 4 groups, based on the type of platelet concentrates used for transfusion.

Results: Patients, were transfused with total number of 647 PC units (235 units were non-leukodepleted and 412 units were leukodepleted). Mean number of PC transfusions per patient was 8.3 PC units, and 4.8 PC unit per one transfusion episode. Before PCT: platelets values were: $18.1 \times 10^9/L \pm 13.1$, Bt 8.4 ± 6.1 min, and after PCT were $28.2 \times 10^9/L \pm 22.1$, 4.7 ± 4.4 min respectively ($p < 0.01$). Mean CCI value was 13.8 ± 30.4 . CCI was corrected in 196/129 PCT and Bt in 122/129 PCT. After supportive therapy using PCs Bt was corrected and became similar in all 4 groups.

Discussion: Clinical output is the most important parameter for treatment decision because many patients can tolerate prolonged periods of profound thrombocytopenia without serious bleeding problems.

Conclusion: In all 4 investigated groups of patients bleeding time was a far better parameter compared with CCI for the PC therapy efficiency.

Authors suggest to be careful and follow clinical and laboratory results personalised to single patient. There is a need to develop better therapies and guidelines so the

Corresponding author:

Primarius Ljubinka I. Nikolić MD, MS

Specialist in Transfusiology

Clinic for Gynecology and Obstetrics, Clinical Centre of Serbia, Višegradska 26, Belgrade, Serbia

E-mail: ljubinka3.9nikolic@gmail.com

practice of platelet therapy can be expected to improve in the future.

Keywords: thrombocytopenia, cancer, platelet transfusion, bleeding

INTRODUCTION

Serious adverse event of anti cancer chemotherapy is granulocytopenia and thrombocytopenia which can decrease efficiency of final therapy results.

The main complications of granulocytopenia is neutropenic sepsis, usually designed as febrile neutropenia. The consequences of leukopenia can be prevented in part of hospitalization in bacteriologically protected units and sterile box and prophylactic usage of granulocyte-colony stimulating factor (G-CSF) and granulocyte-macrophage colony stimulating factor (GM-CSF). Treatment includes a lot of available G-CSF and the CSFs.

Beside there are no such preventive measures and treatment facilities for thrombocytopenia which can result in life-threatening haemorrhagic syndrome, and platelet transfusion remains the only effective approach. Although both autologous bone marrow transplantation and peripheral blood stem rescue have been advocated for high dose chemotherapy haematopoietic support, there is no doubt for importance platelet transfusions in the management of chemotherapy-related haematological toxicities in cancer patients. Advances in platelet transfusion have contributed to improved outcomes in the treatment of cancer patients, allowing high dose chemotherapy delivery with increased safety [1,2].

Continuous supportive platelet therapy is enabled by technological advancement and increased quality of plastic blood bags adjusted for prolonged platelet storage. However, concentrate-specific parameters can be influenced by preparation technique and storage procedures. The duration of storage causes increase of TGF level, which induces an impressive decrease in the in-vivo platelet recovery and survive [3,4,5,6].

The benefits of prophylactic platelet transfusion are controversial and the circulating platelet level which predisposes to haemorrhage is uncertain; so is the effectiveness of alternatives to platelet therapy. Finally, the merits and drawbacks of treatment of thrombocytopenic patients with platelets from single donors and multiple random donors, need

evaluations [7,8].

Several serious scientists through the world after many years are still researching problem of platelet transfusion, but there is no for sure about standpoint [9,10,11].

It has been shown by necessity that many patients can tolerate prolonged periods profound thrombocytopenia without serious problems. A two-fold (double) prolongation of the normal bleeding time, has been reported to be indicative of significant risk of bleeding [12].

THE AIM

To determine whether there is a difference in the clinical efficiency in the use of 4 types of platelet applied for transfusion; - to ascertain whether platelet count increase expressed as corrected count increment (CCI), is a better parameter for the evaluation of platelet transfusion efficiency than the bleeding time (Bt), as the only readily assessable in vivo platelet function related parameter.

SUBJECTS AND METHODS

This paper is a part of academic (noncommercial) IV phase observational nonintervention study, done at National Institute of Oncology and Radiology of Serbia (NIORS), Belgrade. All patients signed informed consent.

Because of technical problems this study was frozen, and this year problem was recognized again so these results are the first step of continuing investigating platelets characteristics.

Clinical bleeding during chemotherapy-induced thrombocytopenia is a serious clinical problem. During several decades this serious cytostatic adverse event is not solved and there is no doctrinal statement about this.

Cancer as itself disease is characterised with non stable hemostatic parameters [13], so it is case to case clinical situation.

Investigation included 78 patients (34 males and 44 females), median age 57 years (range 23-73), diagnosed with lymphoma and metastatic solid tumors, transfused by platelet

concentrates from January till May 1998. None of the patients had disseminated intravascular coagulation (DIC), wide spread adenocarcinoma, infection, or other special situation or diagnosis that might have influenced the outcome of platelet support.

The standards for research laboratories procedures, and investigator were met the required criteria.

Patients were divided into 4 groups, based on the type of platelet concentrates used for transfusion, and produced in Serbian National Blood Transfusion Institute (SNBTI). Four types of platelet concentrates were used depending on Institute supply by the producer.

Group A – patients transfused with leucode-

pleted platelet concentrates (PC) pool obtained from 5 platelet rich plasma PC units (PRP PC) ABO identical, or from 5 buffy coat PC units (BC PC) ABO identical (Leukotrap pooling system*, Cutter USA)

Group B – patients transfused with prestorage filtered PRP PC or BCPC using fourth generation filters (Terumo, Japan)

Group C – patients transfused with standard non-leukodepleted PC (PRP-PC stored 0-5 days) (plastic bags Terumo, Japan) on horizontal shaker at 22°C

Group D – patients transfused with prestorage leukodepleted PC obtained from random donors BC, using spontaneous sedimentation of vertically positioned and stored BCs for 12 hours at 22°C.

Patients group	Type of PC	Lymphoma	MST	N
A	Leukodepleted PC pool	3	24	27
B	Leukodepleted prestorage filtered PC	1	6	7
C	Non-leukodepleted PC from random donors	5	28	33
D	Leukodepleted PC by spontaneous sedimentation	-	11	11
Total		9	69	78

In group A there were 3 patients with lymphoma and 24 with metastatic solid tumors; In group B there was one patient with lymphoma and 6 patients with metastatic solid tumors; In group C there were 5 patients with lymphoma and 28 with metastatic solid tumors and, group D there were 11 patients with metastatic solid tumors. Thus, there were 9 patients with lymphoma and 69 with metastatic solid tumors. Thrombocytopenia in all those patients was serious adverse event (SAE), as consequence of needed aggressive chemotherapy.

Indication for platelet transfusion treatment were:

1. bleeding time prolonged in 108/129 transfusion (Bt 3.4 min – 37 min);
2. when clinical manifestation of haemorrhagic syndrome there present in 2/129 transfusions;
3. when bleeding time was normal but platelets already with WHO grade 4 toxicity range, tested daily expressed tendency towards decrease in 19/129 transfusions.

Out of 108 transfusions in patients with prolonged bleeding time, thrombocytopenia was grade 4 in 97 patients, grade 3 in 8 cases, grade 2 in 2 cases and grade 1 in 1 case.

None of patients had DIC. Grading of thrombocytopenia was performed according to CTC criteria [14].

Efficiency of supportive therapy using PC transfusions was estimated according both to CCI and Duke test for bleeding time (BT) (reference values 1 - 3 minutes), not traumatic

Transfusion indication	No of pts with prolonged BT	No of platelet transfusions
Prolonged bleeding time	108	129
Clinical bleeding	2	129
Decreased Tendency of plt	19	129

as Ivy test. BT and platelet count there were controlled before PC therapy and 20-24 hours after PC transfusion. BT was performed by the same technician although not highly precise technique, was chosen as the only test which evaluates in vivo platelet haemostatic function [15]. Tests were performed 20-24 hours after transfusion since platelet functions are restored during that interval, reflecting clinical effect/benefit of a platelet transfusion. Our presumption was that this could be achieved only by using an in vivo platelet function test or by observing disappearance of clinical manifestations typical for thrombocytopenia.

Platelets were determined using automatic counter Cell Dyn 3500 (Abbott)

Table 1. Patients groups (A,B,C,D) according to transfused platelets

PC-platelet concentrate, MST-metastatic solid tumor

Table 2. Indication for platelet transfusion

obtained from blood specimens using the EDTA. Platelet values were repeated using the sodium citrate and the counting was carried out microscopically in chamber with the 1% ammonium oxalate.

Platelet count was not tested in our patients 1 h after PC transfusion because in that interval only data concerning total number of transfused platelet particles can be obtained. Storage platelets lose viability, decrease 2,3-diphosphoglycerate (2,3 DPG), but can be acceptable as compromise during storage [16]. Around 20-24 hours after transfusion, removal of damaged platelets occurs, and energetic balance of platelets which have lost their energetic balance during storage is restored.

CCI was determined using the following formula:

$$CCI = ((\text{post transfusion platelet count (x10}^9/\text{L)} - \text{pretransfusion platelet count (m}^2\text{)}) / \text{total number platelets transfused})$$

It is generally accepted that an effective PC treatment corresponds to a CCI after 24 h which is ≥ 4.5 ; for the purpose of this study we have considered as effective any CCI over zero (>0) i.e. any platelet increase its relation to pretransfusion one. A corrected bleeding time was considered any shortening below 6 minutes if initial bleeding time was over 6

minutes; if initial bleeding time was 6 minutes or below any shortening was considered as evidence for a corrected bleeding time, meaning that clinical endpoints (i.e. bleeding) are the most important method for evaluating effectiveness of PLT transfusions [17].

In addition on PC investigated patients were transfused with red blood cells (RBC) (concentrates), fresh frozen plasma (FFP) and cryoprecipitate (Cryo): in group A were transfused with RBC and 2 with FFP; in group B, 2 patients were transfused with RBC; in group C, 10 patients were transfused with RBC; One of those 10 patients also receive cryo and another received RBC and FFP. In group D two patients RBC.

Statistical methods used in the investigation were Kruskal-Wallis, Mann Whitney and Wilcoxon Rank Sum W test.

RESULTS

The patient's characteristics are presented on table 1. Out of 78 patients, 33 (41%) were transfused with non-leukodepleted PC, and remaining 45 (59%) with leukocyte depleted PC. Total number of 647 PC units were used. Out of 647 PC units, 235 units (36.3%) were non-leukodepleted and 412 (63.7%) units were leukodepleted. Patients were transfused with the total number of 129 PC transfusions. Mean number of PC transfusions per patient was 8.3 PC units, and 4.8 PC unit per one transfusion (table 1).

The mean platelet count in the investigated patients prior to transfusion was $18.14 \pm 13.1 \times 10^9/\text{L}$, median=15.8; range 1.6-80.0 $\times 10^9/\text{L}$. Platelet levels were: Group A $14.8 \pm 8.1 \times 10^9/\text{L}$; Group B $23.1 \pm 11.6 \times 10^9/\text{L}$; Group C $18.9 \pm 11.8 \times 10^9/\text{L}$; Group D $26.7 \pm 24.9 \times 10^9/\text{L}$. According to platelet count, investigated groups were homogenous before PC transfusions ($p > 0.05$), while according to the BT before PC transfusion, those 4 groups were not homogenous ($p < 0.05$). Prior to transfusions BT was 8.4 ± 6.1 minute for all investigated patients. In group A, BT was 9.1 ± 6.4 minutes, in group B BT was 5.1 ± 1.8 minutes, in group C BT was 9.9 ± 6.3 and in group D BT was 4.2 ± 3.2 minutes.

After PC transfusions, platelet count increase significantly in the total of all investigated patients:

Group A: Results are shown in table 7. Platelet count was noted in a significantly lesser

Table 3. Main patient characteristics and diagnosis

Pts-patients, PC-platelet concentrates, Group A-leukodepleted pool of 6 random donor platelets, Group B-prestorage filtered PC, Group C-nonleukodepleted PC from random donor platelets, Group D-PC leukodepleted after spontaneous sedimentation.

Characteristics	N (%)
Total number of patients	78 (100)
Age (years)	
Median	57
Range	23-73
Sex	
Male	34 (44)
Female	44 (56)
Diagnosis	
Malignant lymphoma	9 (12)
Metastatic solid tumors	69 (88)
Group of pts according to type of PC	
Group A	27 (34)
Group B	7 (9)
Group C	33 (43)
Group D	11 (14)
Number of pts transfused with	
Non leukodepleted PC	33 (41)
Leukodepleted PC	45 (59)
Total number of PC units	647 (100)
Non leukodepleted PC	235 (36.3)
Leukodepleted PC	412 (63.7)

Group	PLT Pretransfusins x10 ⁹ /L	PLT Posttransfusins x10 ⁹ /L	CCI Corrected Count Increment	P value
A	14.8±8.1	22.7±19.3	13.4±34.8	P<0.05
B	23.1±11.6	21.9±7.7	8.6±6.0	p> 0.05
C	18.9±11.8	37.0±26.7	16.0±30.0	p> 0.05
D	26.7±24.9	27.4±15.5	9.4±16.8	p> 0.05
ALL groups	18.14±13.1	28.2±22.1	13.8±30.4	P<0.01

Table 4. Platelet counts before and after PC transfusion

number of performed PC transfusions, 39/53 (73.6%) in relation to the number of corrected Bt, 49/53 (92.4%) PC transfusions (P<0.001). Satisfactory efficiency of platelet transfusions, based either on CCI or BT was noted in 51/53 PC transfusions, i.e. in 96.2% of transfusions. In 14 transfusions (11 patients), there was no expected platelet count increase. Bt was cor-

rected in 12 of those transfusions, and in 2 transfusions, neither platelet count increase occurred, nor the correction of Bt. One of those 2 patients was female with malignant schwannoma and another was a male patient with non-Hodgkin lymphoma. Both patients had a satisfactory platelet count increase CCI and corrected Bt after the second PC transfu-

Group	BT Pretransfusins (minutes)	BT Posttransfusins (minutes)	P value
A	9.1±6.4	4.9±3.7	P<0.01
B	5.1±1.8	3.3±1.9	P<0.05
C	9.9±6.3	5.4±6.0	P<0.01
D	4.2±3.2	2.7±1.9	P<0.01
All groups	8.4±6.1	4.7±4.4	P<0.01

Table 5. Bleeding time before and after platelet transfusion

sions. Patient with malignant schwannoma was transfused with PC on 4 occasions and only one of those transfusions she had unsatisfactory therapeutic response. This patient received no other blood products. Patient with non-Hodgkin lymphoma was transfused with PC on two occasions and after the second transfusion he achieved a positive therapeutic response and received no other blood products. In one female patient urticaria was noted. This patient was transfused only with

after which Bt was corrected, but not platelet count. Two days later, this patient included into group D and was transfused, preceded by that appropriate premedication, and showed satisfactory therapeutic response, both according to CCI and Bt (table 7).

Group B: In two female patients, platelet count was not increased, i.e. satisfactory CCI was not obtained, but Bt was corrected. One patient suffered from malignant schwannoma and the other one from ovarian cancer. None of them

	Group of leukodeplete n=45/78*	Group of non-leukodepleted n=33/51	Total n=78/129
Platelets prior to transfusion			
X±SD(x10 ⁹ /L)	17.76± 13.8	18.9±11.8	18.1±13.1
Median (range)	16.0 (1.8-80)	15.4 (3.7-54.8)	15.8 (1.6-80)
Platelets post transfusion			
X±SD(x10 ⁹ /L)	23.4±17.5	37.0±26.7	28.2±22.1
Median (range)	18.8 (2.7-96.7)	29.0 (3.4-109)	21.8(2.7-109)
CCI (X±SD)	12.2±30.7	16.0±30.0	13.8±30.4
Bt prior to transfusion			
X±SD(min)	7.7±5.9	9.9±6.3	8.4±6.1
Median (range)	7.0 (0.75-32)	8.8 (3.4-37)	7.4 (0.8-32)
Bt post transfusion			
X±SD(min)	4.4±3.4	5.4±6.0	4.7±4.4
Median (range)	3.5 (0.7-14-5)	3.3 (1-32)	3.4 (0.7-32)
Corrected CCI (%)	59/78 (75.6)	47/51 (92.1)	106/129(82.1)
Corrected Bt (%)	73/78 (93.6)	49/51 (96.1)	122/129(94.5)

Table 6. Analysis of platelet transfusion efficiency (leukodepleted versus non-leukodepleted PC)

Table 7. Analysis of Platelet transfusion Efficiency

* No of patients/no of plt transfusions, BT-bleeding time, PC-platelet concentrate

	Group A n=27/53*	Group B n=7/10*	Group C n=33/51*	Group D n=11/15*	Total n=78/129*
Platelets prior to transfusion X±SD(x10 ⁹ /L)	14.8±8.1	23.1±11.6	18.9±11.8	26.7±24.9	18.1±13.1
Median (range)	15.2 (1.8-43.4)	19.8(13.0-39.8)	15.4(3.7-54.8)	17.2(3.9-80)	15.8 (1.6-80)
Platelets post transfusion X±SD(x10 ⁹ /L)	22.7±19.3	21.9±7.7	37.0±26.7	27.4±15.5	28.2±22.1
Median (range)	16.1 (2.7-96.7)	21.1 (11.9-32.7)	29.0 (3.4-109)	22.7 (6.2-53.1)	21.8(2.7-109)
CCI (X±SD)	13.4±34.8	8.6±6.0	16.0±30.0	9.4±16.8	13.8±30.4
Bt prior to transfusion X±SD(min)	9.1±6.4	5.1±1.8	9.9±6.3	4.2±3.2	8.4±6.1
Median (range)	8.2 (0.8-32)	4.9 (2.2-7.3)	8.8 (3.4-37)	3.4 (1-10)	7.4 (0.8-32)
Bt post transfusion X±SD(min)	4.9±3.7	3.3±1.9	5.4±6.0	2.7±1.9	4.7±4.4
Median (range)	3.8 (7-14.5)	3.3 (0.7-5.8)	3.3 (1-32)	1.8 (0.7-6.3)	3.4 (0.7-32)
Corrected CCI (%)	39/53(73.6)	8/10(80)	47/51(92.1)	12/15(80)	106/129(82.1)
Corrected Bt (%)	49/53(92.4)	10/10(100)	49/51 (96.1)	14/15(93.3)	122/129(94.5)
No of uncorrected CCI	14	2	4	3	23
No of uncorrected CCI and corrected BT	12	2	4	2	20
No of uncorrected BT	2	0	2	1	5
No of uncorrected CCI& BT	2	0	0	1	3
No of uncorrected BT and corrected CCI	0	0	2	0	2
No of uncorrected CCI and/or BT	14	2	6	3	25
Transfusion adverse reaction - urticarial	1	0	0	0	1
No of pts transfused with FFP	2	0	1	0	3
No of pts transfused with Cryoprecipitate	0	0	1	0	1
No of pts transfused with RBC	4	2	11	2	21
No of PC units per group	263	57	235	92	647
No of transfusions	53	10	51	15	129
Mean No of PC units per transfusion	4.8±1.1	5,6±1.1	4.6±1.6	6.6±1.4	5.0±1.4
Mean No of transfusion per patient	1.96	1.43	1.54	1.36	1.65
Mean No of PC units per patient	9.3	8.1	7.1	8.4	8.3

received any other blood products. Based on CCI, 8/10 PC transfusions, and according to Bt, all showed satisfactory response.

Group C: In 4 PC transfusions (4 patients) was not corrected 20-24 hours after the procedure, while platelet count had increased. CCI was satisfactory in 47/52 (92.1%) and Bt was corrected in 49/51 (96.1%) PC transfusions. According to CCI and/or Bt, PC transfusions gave satisfactory result in 51/51 (100%) transfusions in group C. Bt before PC transfusion was 9.9 ± 6.3 min, median=8.8, range 3.4-37 minutes. Bt was not corrected in one female patient with NHL and in one patient with testicular cancer. CCI was not satisfactory in the patient with NHL, who had increase platelet count and uncorrected Bt in previous PC transfusion, as well as in the patient with metastatic solid tumors (leiomyosarcoma of the uterus, in one with gastric cancer, and in one with ovarian cancer). In this group there were no immediate transfusions reactions (table 7).

Group D: In three PC transfusions given to 3 patients CCI was not corrected. In 2 of these 3 patients Bt was corrected. One of those patients suffered from laryngeal cancer and another one had malignant schwannoma (who was transfused one month earlier in group one with leukodepleted PC). Out of these 3 patients, in one patient (female) with metastatic cancer with unknown primary tumor localisation, neither platelet count was increased nor Bt corrected. Three days later, the patient was transfused with the same dose of pooled leukocyte depleted platelets with a satisfactory clinical haemostatic effect, platelet count increase and corrected Bt. Unsatisfactory response in the first use of PC remains unexplained. Based on CCI, PC transfusions were efficient in 12/15 (80%) PC transfusions, and according to Bt in 14/15 (93.3%) transfusions (table 7).

Platelet count value before PC transfusion for the group of patients who were transfused with leukocyte depleted platelets (groups A+B+D) was $17.76 \pm 13.79 \times 10^9/L$, median=16.0, range 1.8-80 $\times 10^9/L$. Bt before PC transfusion was 7.66 ± 5.93 min, median=7.0, range 0.75-32 minutes. After PC transfusion, platelet count was $23.40 \pm 17.48 \times 10^9/L$, median=18.80 range 2.67-96.7. Bt after PC transfusion was 4.38 ± 3.42 minutes, median=3-54, range 0.67-14.5. CCI for groups A+B+D was 12.17 ± 30.73 , median=6.82, range 0.26-76.20. Platelet count increase i.e. positive

CCI was found in 59/78 (75.6) transfusions and corrected Bt in 73/78 (93.6%) (table 7).

In total, a corrected CCI was found for 106/129 transfusions (82.1%) and corrected Bt for 122/129 (94.5%) PC transfusions ($p=0.002$ t.j. $p<0.01$).

DISCUSSION

The investigated 4 groups were homogenous before platelet transfusions according to platelet count ($p=0.4$) and not homogenous according to Bt ($p<0.01$ t.j. $p=0.006$). After supportive therapy using PCs Bt was corrected and became similar in all 4 groups. Nonhomogeneity among 4 groups disappeared ($p=0.44$) which possibly pointed to the positive effects of supportive platelet therapy and successful therapeutic result. These results correlated with literature data [3,7]

Concerning platelet count and Bt of all investigated patient groups there were no significant correlation either before ($\rho=-0.08$) or after PC transfusions ($\rho=-0.16$) [9].

Significant platelet increase was noted in group A ($p<0.05$ t.j. $p=0.046$) and group C ($p=0.0001$), in patients transfused with leukodepleted platelets (groups A+B+D) ($p<0.05$ $p=0.0239$), as well as in all patients together (groups A+B+C+D) ($p<0.00001$). In groups B and D the number of patients was small so that significance could not be determined, although the increase was noted.

Compared with group A, group C had significantly higher platelet count after PC transfusion ($p=0.0048$), as well as a higher CCI compared with group A ($p=0.0248$). According to post transfusion Bt, groups C and A did not differ significantly ($p=0.9694$).

Groups treated with leukocyte depleted PC (groups A, B and D) and group treated with non-leukocyte depleted PC showed homogeneity before transfusion both according to platelet count ($p=0.8$) and Bt ($p=0.07$).

Non-leukodepleted PC treated group (group C) compared with leukodepleted PC treated group (A+B+D) showed significantly higher platelet count increase ($p<0.01$ t.j. $p=0.0082$), and significantly higher CCI ($p<0.05$ $p=0.0139$) after PC transfusion. Higher efficiency of non-leukodepleted PC could be explained partly by the presence of plasma and leukocyte contents (PGE1, IL6) [18,19,20,10], and partly by the loss of a

population of platelets during leukodepletion procedure (in groups A, B, D) and the loss of cytokines which take part in platelet function and viability maintenance. However non-leukodepleted platelets are not recommended if multiple platelet transfusion are required, in order to decrease the exposure to alloantigens, i.e. in order to postpone alloimmunization and refractorines [21,22,23,24,11].

Despite of non-uniforme numeric results and wide differences in types of malignant diseases, clinically transfusion therapy outcome was satisfactory which is in correlation literatures dates. After more than ten years there is still dilemmas and little bit confusion related to recommended values of platelet increase, shortening bleeding time and clinical outcome [9,13,21].

Bt after PC transfusion showed no statistical difference between the groups of leukodepleted and non-leukodepleted PC treated patients ($p=0.61$). Leukocyte presence in group C did not induce occurrence of posttransfusion reactions. One posttransfusion reaction occurred in group A and could be explained by the reaction to plasma present in platelet supernatant [21].

BT were significantly corrected in all patient groups in relation with the BT prior to platelet transfusion ($p<0.01$).

In 23 PC transfusions platelet count and CCI increase did not occur. Out of 23 PC transfusions with unsatisfactory CCI, 5 were performed in 5 males, and 18 in 13 female patients. None of the female patients was treated by any other blood product. Likewise, the only posttransfusion reaction occurred in a female patient. As all these females were uni or multiparous this could speculatively be explained by the contact with another antigenic system during pregnancy [25,26].

In 20 out of 23 PC transfusions where platelet increase did not occur and satisfactory CCI was not achieved, BT was corrected. In all 4 investigated groups BT was corrected in significantly higher percentage of PC transfusions compared with CCI ($p=0.002$).

To improve efficiency of platelet transfusion in patients who develop refractoriness, and to prevent or delay the onset of refractoriness, the storage time should be shortened [3,27,11]; single donor platelets (SD PC) should be used whenever possible rather than standard PC [28,11]. Transfusion of high platelet doses increases the transfusion inter-

val and can reduce the number of Platelet concentrates required by thrombocytopenic patients that significantly reducing donor exposure [29,11]. HLA an HPA-matched platelets [30,31]; or as an alternative HLA class I eluted platelets should be used for successful treatment of platelet transfusion refractoriness [32]. Washed platelet concentrates in patients with febrile non-haemolytic transfusion reactions should be used [33]. Autologous platelet transfusions are the best choice [34,11].

Disadvantages using prophylactic PC therapy include increased number of donor exposures and possibly also enhanced risk of alloimmunization and refractorines [35,36,10,11]. The advantages of prophylactic therapy are still not universally accepted.

Even when the value of prophylactic therapy is accepted the platelet count threshold at which this is justified is still very much a matter of debate and it is certainly true that not all patients with severe thrombocytopenia should be considered automatic candidates for prophylactic therapy [37,38,39].

Since, many patients can tolerate prolonged periods of profound thrombocytopenia without serious bleeding problems [10,11], bleeding time is much better parameter to assess (for assessment) the need for platelet transfusions than platelet level.

Platelet transfusion practice is being questioned more than ever before. As we develop better therapies and guidelines, the practice of platelet therapy can be expected to change in the near future [10, 11].

CONCLUSION

The advantages of prophylactic PC therapy are not still universally accepted, but authors agree with other published papers, that prophylactic platelet transfusion is clinically recommended.

Advantages using prophylactic PC therapy include expenses preventing further costs of bleeding complications. Disadvantages are increased number of donor exposures and possibly also enhanced risk of alloimmunization.

These data show that beside PC therapy efficiency monitoring, which includes platelet counts and CCI, Bt in vivo platelet function test should also be performed. Since bleeding time was corrected in cases of uncorrected CCI, uncorrected CCI should not be considered as refractorines to PC without the deter-

mination of bleeding time. PC transfusions followed by a satisfactory CCI but uncorrected Bt associated with clinical improvement failed should not be considered as successful ones.

In all 4 investigated groups of patients bleeding time was a far better parameter compared with CCI for the PC therapy efficiency. Bleeding time is much better parameter than platelet level for assessment the need for platelet transfusions.

Considering to cancer disease etiology, course of disease, and patient clinical status in this sophisticated part of treatment success, it is necessary to assess clinical cost benefit results in any patient separately.

Authors suggest to be careful and follow clinical and laboratory results personalized to single patient.

There is a need to develop better therapies and guidelines so the practice of platelet therapy can be expected to improve in the future. In the next part of this investigation we shall underline personalized medical treatment.

ACKNOWLEDGMENTS

Irena Vukajlović for technical assistance.

REFERENCES

1. Sarkodee-Adoo C, Schiffer CA Platelet transfusion support for patients with cancer and hematologic malignancies, *Curr Opin Hematol* (1996). 3(5):347-354
2. Hashiguchi Y, Fukuda T, Yoshida H, Ichimura T, Matsumoto Y, Yasui T, Sumi T, Ishiko O. Platelet transfusion during chemotherapy-induced thrombocytopenia in patients with gynecologic malignancy. *Ann Oncol* (2013) 24 (suppl 9): ix84.doi: 10.1093/annonc/mdt460.91 11th Annual Meeting of the Japanese Society of Medical Oncology, 29-31 August 2013, Sendai, Japan. Abstract P2
3. Bock M., Muggenthaler K.H., Schmidt U., Heim M.U., Mempel W (1995). Post-transfusion rise in thrombocytes: observations in a hematologic-oncologic patient sample, *Infusions ther Transfusions Med* 22(6): 350-354
4. Rebull P. Platelet Refractoriness. *European Hematology Review*, 2007;1(1):21-2
5. Goodnough, L.T., Riddell, J., Lazarus, H. et al. Prevalence of platelet transfusion reactions before and after implementation of leukocyte-depleted platelet concentrates by filtration. *Vox Sang*.1993; 65: 103-107
6. Goodnough, L. T., Maggio, P., Hadhazy, E., Shieh, L., Hernandez-Boussard, T., Khari, P. and Shah, N. Restrictive blood transfusion practices are associated with improved patient outcomes. *Transfusion*, 2014; 54: 2753-2759.
7. Rebull P. Platelet transfusion trigger in difficult patients. *Transfus Clin Biol*. 2001 Jun; 8(3):249-54.
8. Stanworth SJ. A No-Prophylaxis Platelet-Transfusion Strategy for Hematologic Cancers. *The new England journal of medicine*, may 9, 2013 vol. 368 no. 19
9. Vamvakas EC. Evidence-based practice of transfusion medicine: is it possible and what do the words mean? *Transfus Med Rev*. 2004; 18(4):267-78.
10. Vamvakas EC. Allogeneic blood transfusion and cancer recurrence: 20 years later. *Transfusion*. 2014; 54(9):2149-53.
11. Annen K, Olson JE. Optimizing platelet transfusions. *Curr Opin Hematol*. 2015 Nov; 22(6):559-64.
12. Schiffer CA. Hematological cancer: Prophylactic platelet transfusion is frequently not necessary *Nature Reviews Clinical Oncology* 2013(10): 431-432
13. Djukić VB, Kastratović DA, Pendjer IP, Majstorović BM, Nikolić Lj, Borčić IV, Vujčić ZN. Patient with double cancer--successfully treated. *Acta Chir Iugosl*. 2005;52(3):91-3
14. Common toxicity Criteria. In: Investigator's Handbook. A manual for participants in clinical trials of investigational agents sponsored by the Division of Cancer Treatment, National Cancer Institute. NIH Pub 1993. 93-2770
15. Brinkhous KM. W. W. Duke and His Bleeding Time Test A Commentary on Platelet Function. *JAMA*. 1983; 250(9):1210-1214.
16. Thibault L, Beauséjour A, de Grandmont MJ, Lemieux R, Leblanc JF. Characterization of blood components prepared from whole-blood donations after a 24-hour hold with the platelet-rich plasma method *Transfusion* 2006 Aug;46(8):1292-9
17. Holbroa A, Infantia L, Sigleb J, Busera A. Platelet transfusion: basic aspect. *Swiss med Wkly*, 2013;143:w13885
18. Bishop J.F., Matthews J.P., Yaen K., McGrath K The definition of refractoriness to platelet transfusions. *Transfusion Medicine*. 1992;2:35-41
19. The Trial to Reduce Alloimmunization to Platelets Study Group (1997). Leukocyte reduction and ultraviolet B irradiation of platelets to prevent alloimmunization and refractoriness to platelet transfusions. *N Engl J Med* 1997; 337(26):1861-1869
20. Locker GJ, Staudinger T, Knapp S, Laczika KF, Burgmann H, Urllicic A, Wgner A, Metnitz P, Rnoebl P, Schuster E, Frass M. Prostaglandin E1 inhibits platelet decrease after massive blood transfusions during major surgery: influence on coagulation cascade?, *J Trauma* 1997. 42(3):525- 531

21. Heddle NM, Klama L, Singer J, Richards C, Fedak P, Walker I, Kelton JG The role of the plasma from platelet concentrates in transfusion reactions, *N Engl J Med*. 1994.331(10):625-628
22. Ishida A, Handa M The efficacy of leukodepleted platelet transfusion, *Nippon Rinsho*. 1997, 55(9):2385-2391
23. Kunz D, Luley C, Heim MU, Bock M. Transforming growth factor beta is increased in plasma of patients with hematologic malignancies after transfusion of platelet concentrates, *Transfusion* 1998;38(2): 156-159
24. Seftel MD, Grove GH, Petraszko T, Benny WB, Le A, Lee CY, Spinelli JJ, Sutherland HJ, Tsang P, Hogge DE. Universal prestorage leukoreduction in Canada decreases platelet alloimmunization and refractoriness. *Blood*. 2004;103 (1): 333-339
25. Mollison PL, Engelfriet CP, Contreras M. The transfusion of platelets, leucocytes, hematopoietic cells and plasma components. In: *Mollison's Blood Transfusion in Clinical Medicine*. Oxford, Blackwell Scientific Publications, UK 11th edition, chapter 14; 2005: 611-665
26. Napier J.A.F. *Handbook of Blood Transfusion Therapy*. John Wiley&Sons, Chichester, second edition, 1995: 75-94.
27. Duguid JK, Carr R, Jenkins JA, Hutton JL, Lucas GF, Davies JM. Clinical evaluation of the effects of storage time and irradiation on transfused platelets. *Vox Sang*. 1991; 60(3):151-4.
28. Anderson NA., Gray S., Copplestone JA., Chan DC., Hamon M., Prentice AG., Johnson SA., Philips M., van Waeg G., Oakhill A., Abeyasekera S., Pamphilon DH A prospective randomized study of three types of platelet concentrates in patients with hematological malignancy: corrected platelet count increments and frequency of nonhemolytic febrile transfusion reactions, *Transfus Med*, 1997;7(1):33-39
29. Norol F, Bierling P, Roudot-Thoraval F, Le Coeur FF, Rieux C, Lavaux A, Kuentz M, Duedari N. Platelet transfusion: a dose-response study, *Blood*. 1998. 92(4): 1448-1453
30. Kekomaki S, Volin L, Koistinen P, Koivunen E, Koskimies S Ruutu T, Timonen T, Kekomaki R. Successful treatment of platelet transfusion refractoriness: the use of platelet transfusions matched for both human leucocyte antigens (HLA) and human platelet alloantigens (HPA) in alloimmunized patients with leukemia, *Eur J Haematol*. 1998. 60(2): 112-118
31. Meinke S, Sandgren P, Mörtberg A, Karlström C, Kadri N, Wikman A, Höglund P. Platelets made HLA deficient by acid treatment aggregate normally and escape destruction by complement and phagocytes in the presence of HLA antibodies. *Transfusion*. 2015 Oct 7. doi: 10.1111/trf.13350. [Epub ahead of print]
32. Novotny VM, Huizinga TW, van Doom R, Briet E, Brand A. HLA class I-eluted platelets as an alternative to HLA-matched platelets, *Transfusion* 1996. 36(5):438-444
33. MacWhannell A, Smith N, Thomas J (1999). The use of Washed Platelet Concentrates in Patients with febrile non-hemolytic transfusion Reactions, 11th MASCC International Symposium Supportive Care in Cancer Feb. 18-20, 150
34. Pedrazzoli P, Perotti C, Noris P, Da Prada GA, Zibera C, Battaglia M. Gibelli N, Preti P. Pavesi L, Torretta L, Balduini CL, Salvaneschi L, della Cuna GR. Autologous platelet transfusion in patients receiving high-dose chemotherapy and circulating progenitor cell transplantation for stage II/III breast cancer, *Haematologica* 1998. 83(8):718-23
35. Roy, A.J., Jaffe, N. and Djerassi, I. Prophylactic platelet transfusions in children with acute leukemia: a dose response study. *Transfusion*. 1973;13:283-290
36. Gmur, J., Burger, J., Schanz, U., Fehr, J. and Schaffner, A. Safety of stringent prophylactic platelet transfusion policy for patients with acute leukemia. *Lancet*. 1991. 338:1223-1226.
37. Seon Young Kim, Ji-Eun Kim, Hyun Kyung Kim, Kyou-Sup Han, Cheng Hock Toh. Accuracy of Platelet Counting by Automated Hematologic Analyzers in Acute Leukemia and Disseminated Intravascular Coagulation: Potential Effects of Platelet Activation. *Am J Clin Pathol*. 2010; 134(4):634-647.
38. Kunal Sehgal, Y. Badrinath, Prashant Tembhare, P. G. Subramanian, Sanjay Talole, Ashok Kumar, Vijaya Gadage, Shashikant Mahadik, Sitaram Ghogale, Sumeet Gujral. Comparison of Platelet Counts by CellDyn Sapphire (Abbot Diagnostics), LH750 (Beckman Coulter), ReaPanThrombo Immunoplatelet Method (ReaMetrix), and the International Flow Reference Method, in Thrombocytopenic Blood Samples. *Cytometry Part B (Clinical Cytometry)*.2010; 78: 279-285.
39. Mishima Y, Tsuno NH, Matsuhashi M, Yoshizato T, Sato T, Ikeda T, et al. Effects of universal vs bedside leukoreductions on the alloimmunization to platelets and the platelet transfusion refractoriness. *Transfus Apher Sci*. 2014 Nov 11. [Medline]

Upotreba četiri vrste koncentrata trombocita u tretmanu trombocitopenije izazvane hemioterapijom

Ljubinka I. Nikolić¹, Ninoslav D. Nedeljković², Svetislav B. Jelić²,
Nada D. Suvajdzic Vuković³, Ivana M. Filipović-Lješković², Srdjan Z. Marković⁴,
Drina Lj. Janković⁵, Dragana A. Kastratović⁴

¹ Klinika za ginekologiju i akušerstvo, Klinički centar Srbije, Beograd, Srbija

² Institut za onkologiju i radiologiju Srbije, Beograd, Srbija

³ Klinika za hematologiju, Klinički centar Srbije, Beograd, Srbija

⁴ Klinički centar Srbije, Beograd, Srbija

⁵ Institut za nuklearne nauke Vinča, Beograd, Srbija

KRATAK SADRŽAJ

Uvod: Ozbiljni neželjeni efekti antikancerske hemioterapije su granulocitopenija i trombocitopenija, koje mogu smanjiti efikasnost u konačnom ishodu terapije.

Nakon više godina transfuzija koncentratima trombocita (PCT) je još uvek predmet istraživanja bez sigurnih preporuka.

Cilj: Determinisati da li postoji razlika u kliničkoj efikasnosti ukoliko se za transfuziju koriste 4 tipa koncentrata trombocita (PC); - proceniti da li je porast broja trombocita, izražen kao korigovani porast broja trombocita (CCI), bolji parametar za procenu efikasnosti transfuzije trombocita od vremena krvarenja (Bt) kao jedinog testa za procenu funkcije trombocita *in vivo*.

Materija i metode: Ovaj rad je deo IV faze akademske (nekomercijalne) opservacione neinterventne studije. U istraživanje je uključeno 78 pacijenata sa dijagnozom malignog limfoma i metastaskim solidnim tumorima koji su transfundovani koncentratima trombocita. Pacijenti su, na osnovu vrste trombocitnih koncentrata korišćenih za transfuziju, podeljeni u 4 grupe.

Rezultati: Za transfuziju ispitivanih pacijenta je korišćeno ukupno 647 jedinica PC od čega je 412 jedinica bilo osiromašeno leukocitima, a 235 nije. Prosečno je korišćeno 8.3 jedinice PC po pacijentu i 4.8 PC jedinica za jednu transfuziju.

Pre PCT: vrednosti trombocita su bile: $18.1 \times 10^9/L \pm 13.1$, Bt 8.4 ± 6.1 min, a nakon transfuzije $28.2 \times 10^9/L \pm 22.1$, 4.7 ± 4.4 min respektivno ($p < 0.01$). Srednja vrednost CCI je bila 13.8 ± 30.4 . CCI je bio korigovan u 196/129 PCT, a Bt u 122/129 PCT. Nakon suportivne terapije koncentratima trombocita vreme krvarenja se korigovalo i nije bilo statistički značajne razlike među grupama ($p > 0.05$).

Diskusija: S obzirom da mnogi pacijenti oboleli od malignih bolesti mogu da tolerišu prolongirane periode izražene trombocitopenije bez pojave ozbiljnih krvarenja, pri donošenju odluke za klinički tretman najznačajniji parametar je klinički ishod.

Zaključci: U sve 4 istraživane grupe pacijenata vreme krvarenja je bilo mnogo bolji parametar od CCI za procenu efikasnosti PC terapije. Autori sugerišu opreznost i redovni monitoring kliničkih i laboratorijskih rezultata u skladu sa postulatima personalizovanog tretmana. Neophodno je razvijati bolje terapijske pristupe i vodiče za kliničku terapijsku upotrebu koncentrata trombocita u neposrednoj budućnosti.

Ključne reči: trombocitopenija, kancer, transfuzija trombocita, krvarenje

Received: October 31, 2015
Accepted: December 3, 2015